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NOTE

THE BASE CATALYZED REARRANGEMENT OF 3-0-DEMETHYLFORTIMICIN A

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Fortimicins A (1) and B (2) are aminoglycoside antibiotics. Both compounds consist of a 6-*epi*purpurosamine B moiety and the aminocyclitol fortamine. Fortimicin A differs from B in having a glycyl substituent at its 4-*N*-position. Base catalyzed loss of glycine from the 4-amino group of 1 to give 2 and a rearrangement product, 2'-*N*-glycylfortimicin B or isofortimicin (3), has been reported by TADANIER, *et al.*¹⁾ In that study, a number of 4-*N*-acylfortimicin analogs

Scheme 1. Some mass spectral fragments of 5 and 6.





Scheme 2. Proposed mechanism for the rearrangement of 4.



	5			6		
	pD 1.16	pD 10.04	β -Shift	pD 2.64	pD 10.89	β -Shift
C-1	53.4	53.1		53.7	53.9	0.2
C-2	71.4	72.9	0.5	70.7	74.8	4.1
C-3	65.8	69.2	3.4	65.0	69.6	4.6
C-4	58.4	64.6	6.2	60.1	65.1	5.0
C-5	65.8	69.7	3.9	67.1	70.9	3.8
C-6	75.3	76.5	0.7	73.7	83.1	9.4
C-1'	95.5	98.8	3.3	97.1	99.1	2.0
C-2'	51.8	51.2	_	52.1	50.7	
C-3'	21.6	26.8	5.2	22.4	23.7	1.3
C-4'	26.3	26.9	0.6	27.1	26.8	
C-5'	70.8	72.0	1.2	68.5	71.6	3.1
C-6'	49.3	50.3	1.0	49.0	50.1	1.1
C-7'	15.2	17.1	1.9	15.1	18.4	3.3
C-2''	41.4	45.2	3.8	41.3	44.8	3.5
$N-CH_3$	31.6	35.3	3.7	32.6	35.5	2.9

Table 1. 25 MHz CMR parameters of compounds 5 and 6.

were shown to undergo similar acyl migrations. SHIRAHATA, *et al.*²⁾ also reported similar rearrangements for fortimicin D (6'-demethylfortimicin A) and some further 4-*N*-acylfortimicin analogs. In both studies, however, the only rearrangement products observed were of acyl migration from the 4-amino to the 2'-amino group. Thus the mechanism for the rearrangement has been postulated to involve intramolecular migration of the 4-*N*-acyl moiety *via* the 5-hydroxy group to the 2'-amino group.

We wish to report that 3-O-demethylfortimicin A (4), a synthetic analog³⁾ of 1, undergoes base catalyzed rearrangement to afford 1-Nglycyl-3-O-demethylfortimicin B (5) and 3-Odemethylisofortimicin (6). Compound 5 has been reported by MCALPINE, *et al.*⁴⁾ as a minor fermentation product (fortimicin AN) of *Micromonospora olivoasterospora*, the producing organism of 1. Compound 6 has not hitherto been reported.

The compounds were obtained by treatment of 4 with 10 \times NH₄OH for 18 hours, the resulting mixture was neutralized and chromatographed over IRC-50 (NH₄⁺) using an aqueous-IM NH₄OH gradient. The fractions collected were neutralized and combined after thin-layer chromatography (t.l.c.) using silica gel and the solvent system; CH₂Cl₂–MeOH–NH₄OH (1:1:1, lower phase). The combined fractions were desalted over Sephadex G-10 and lyophilized to afford 5, 6 and 3-O-demethylfortimicin B (7) in 51, 28 and 19% yields, respectively. Identification of 5 as fortimicin AN was by direct comparison of spectra and tlc characteristics. The product of simple glycine cleavage, compound 7, was similarly shown to be identical with an authentic sample³⁾ of 3-O-demethylfortimicin B. The spectral characteristics that led to the identification of the rearrangement products are the following:

Compound 5; MS (Scheme 1) m/e: 391 (M⁺), 392 (M+H⁺), 374 (M⁺-NH₃), 278 (12), 250 (10), 143 (9); PMR (D₂O) δ ppm from external TMS: 1.5 (3H, d, J=7 Hz, CH₃-7'), 1.7~2.4 (4H, CH₂-3', 4'), 2.9 (3H, S, HNCH₃-4), 3.1~3.4 (2H, CH-2', 6'), 3.4 (1H, CH-4), 3.85 (2H, S, CH₂-2''), 4.0 (1H, CH-5'), 4.2~4.5 (2H, CH-2, 6), 4.6~4.7 (2H, CH-3, 5), 4.8 (1H, HC-NCO-1), 5.8 (1H, d, J=3.5, CH-1'); IR (KBr) cm⁻¹: 1570, 1650 (HN-C=O).

Compound **6**; MS m/e: 391 (M⁺), 392 (M⁺ H⁺), 374 (M⁺ - NH₈), 221 (**13**), 200 (**8**), 193 (**11**); PMR (D₂O) δ ppm from external TMS: 1.7 (3H, d, J=7 Hz, CH₈-7'), 2.1 ~ 2.5 (4H, CH₂-3', 4'), 3.0 (3H, S, HNCH₈-4), 3.4 ~ 3.7 (2H, CH-1, 6'), 3.8 (1H, CH-4), 4.0 (1H, CH-5'), 4.2 (2H, S, CH₂-2''), 4.3 ~ 4.4 (2H, CH-2, 6), 4.5 (1H, CH-2'), 4.6 ~ 4.7 (2H, CH-3, 5); IR (KBr) cm⁻¹: 1570, 1650 (HN-C=O).

The 4-N-CH₃ singlets at δ 2.9 and 3.0 in the PMR spectra of 5 and 6, respectively, indicated that the 4-amino group in each case was unsubstituted by an acyl group, since 4, which has a glycyl substituent on the 4-amino group, shows a chemical shift of δ 3.62 for the 4-N-CH₃ group.³⁾ Hence the point of attachment of glycine was not the 4-N-CH₃ position in either 5 or 6. The position of glycine substitution was determined for each compound by spin decoupling experiments. For compound 5, the C-1 proton was determined to be at δ 4.8 by decoupling the resonances for the C-2 and C-6 protons. Similarly, the C-2' proton was located at δ 4.5 for compound **6** by decoupling the C-1' proton. The chemical shifts for the C-1 and C-2' protons of 5 and 6 have been reported⁵⁾ as 4.0 and 3.3, respectively. The downfield shifts observed for the C-1 proton in 5 and the C-2' proton in 6 indicated the position of glycine substitution to be the 1-NH₂ group in 5 and the 2'-NH₂ group in **6**.

The m/e 250 (10) and 278 (12) fragments (Scheme 1) in the mass spectrum of 5 was consistent with glycine substitution on the 3-Odemethylfortamine cyclitol. On the other hand, the spectrum for 6 showed peaks at m/e 193 and 221, consistent with a fortamine cyclitol lacking a glycyl substituent, and a peak at m/e 200, confirming that glycine was attached to the 6-*epi*purpurosamine B moiety. In addition, 5 showed a peak at m/e 143, consistent with an *epi*-purpurosamine B moiety.

The CMR spectra of **5** and **6** are summarized in Table 1. A comparison of the β -shifts observed for the various carbon assignments in the spectra of the two compounds showed differences in magnitude that were diagnostic. Thus **5**, which is acylated at the 1-NH₂ group showed only small shifts for C-2 and C-6, upon titration, as compared to the same carbons in **6**. Similarly, the shifts for C-1' and C-3' in **6**, which is acylated at the 2'-NH₂ group were smaller than those observed for the same carbons in **5**. The spectral evidence is thus consistent with the assigned structures of the rearrangement products of **4**.

It is of interest that 4-N-acyl fortimicins undergo base catalyzed rearrangements to 2'-N-

acyl derivatives exclusively,^{1,2)} whereas **4**, which differs from the above compounds only in lacking a 3-*O*-methyl group, rearranges to give 1-*N*- and 2'-*N*-acyl products. We suggest an intramolecular mechanism involving the formation of a 5-*O*-acyl derivative **14** (Scheme 2) of the type described by SHIRAHATA, *et al.*²⁾ In this case however, the two conformers, **15** and **16**, may both be stable and undergoing rearrangement. Hence direct migration of glycine from the 5-position of **15** to the 1-*N* affords **5**, whereas migration from 5-*O* to 2'-*N* in **16** affords **6**.

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